

Qualitative Phytochemical Screening and Antibacterial Activity of *Moringa Oleifera* Linn and *Terminalia Arjuna* Linn

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ABSTRACTS

The current examination was completed to decide extractive qualities, explore the phytochemical constituents and antimicrobial activities of two tree species *Moringa oleifera* Linn. and *Terminalia arjuna* Linn. *Terminalia arjuna* has generally been utilized specially to treat coronary illness and *Moringa oleifera* is one of the assortments of customary nutritious species. The extractive value estimations of *Moringa oleifera* and *Terminalia arjuna* were resolved utilizing various solvents, for example water, methanol, ethanol, acetone and chloroform. Phytochemical examination uncovered the nearness of different significant optional metabolites, for example, sterols, terpenoids, alkaloids, sugars, glycosides, proteins, amino acids, steroid, and tannins in different concentrates.

An examination was directed to contemplate the antibacterial activities utilizing methanolic and chloroform concentrates of *Moringa oleifera* and *Terminalia arjuna*. The action was examined utilizing agar well dissemination technique against *Bacillus subtilis*, *Escherichia coli* (NCIM-2066), *Klebsiella pneumoniae* (NCIM-2883), *Shigella dysenteriae* and *Staphylococcus aureus*, which frequently cause enteric diseases in Humans. Remarkably, our studies have demonstrated that *Moringa oleifera* leaves and *Terminalia arjuna* barks extracts possess bactericidal activities against a range of different disease-causing bacterial species in humans such as *E. coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. These two plants have potential to be considered in one of the new infections fighting strategies in controlling bacterial diseases and could be a good source of could be source of new antibiotic compounds.

Keywords: *Moringa oleifera*, *Terminalia arjuna*, Antibacterial, solvent, Extractive values, Phytochemical testing, *Bacillus subtilis*, *Escherichia coli*, *Klebsiella pneumoniae*, *Shigella dysenteriae* and *Staphylococcus aureus*

INTRODUCTION

Medicinal plants have been utilised in conventional medicines for centuries and it has played a crucial role in preventing diseases. Phytochemicals chemical derived from plants and its synthetic constituents are liable for the shading and different properties of the plant. A few phytochemicals have a wide scope of exercises, which confers resistance against long term diseases. The phytochemicals like alkaloids, flavonoids, tannins, saponins, sugars, glycosides, Phyto-sterols, phenols, protein and amino corrosive, diterpenes and so forth are referred to show therapeutic actions along with their physiological activities (Nadkarni et.al. 2009).

Moringa oleifera has a place with the group of Moringaceae, local to India, Africa, Arabia, South Asia, and the pacific and Caribbean Islands. *Moringa oleifera* is a little, quickly developing evergreen or deciduous tree that normally grows up to 9-12 m tallness, open crown of hanging delicate branches, fluffy foliage of outing natural leaves and thick corky, whitish bark (NASIR, 1972; Parrotta, 2001; Ramachandran et.al.,1980). *Moringa oleifera* is normally known as "Drumstick". It is a little or medium estimated tree, found in the sub-Himalayan tract (Gupta RK ,2010). *Moringa* leaves contains phytochemical having strong anticancer, antimicrobial and are viewed as brimming with therapeutic properties (Tiloke et.al.,2013; Caceres, 1991)

Terminalia arjuna (*T. arjuna*) is a deciduous enormous measured fluted tree to 30 m tall and 2-2.5 m breadth at bosom tallness, with a frequently buttressed trunk. *T. arjuna* has a place with the group of Combretaceae, it is a huge tree, found all through the South Asian district. This tree is typically evergreen (Ali, M. 1994). The bark of *T. arjuna* is hostile to dysentery, antipyretic, astringent, cardiotoxic, Lithotriptic, anticoagulant, hypolipidemia, antimicrobial specialist (Phani Kumar, 2013; Ram et al., 1997; Nema et al. 2012). The powder of the bark goes about as a diuretic in cirrhosis of liver and gives alleviation in suggestive hypertension (Chatterjee, A. S., 1994). Antibacterial obstruction has become a worldwide issue. Techniques to improve the momentum circumstance remember inquire about for finding new and creative antibiotics. The chemotherapeutic operators have been of incentive in controlling numerous diseases however they rely upon sensible use to limit the occurrence of creating obstruction. Due to the cost-effective antibacterial properties, a huge extent of the populace uses restorative plants for the treatment of irresistible illnesses. As per World Health Organization (WHO), over 80% of the total population depends on customary medicine (Chatterjee, A. S. 1994; World Health Organization. (2002).

The therapeutic estimation of plants lies in some synthetic substances that produce an unmistakable physiologic activity on human body. The most significant bioactive mixes of plants are alkaloids, flavonoids, tannins and phenolic compounds. The phytochemical investigate dependent on ethno pharmacological data is commonly viewed as a viable methodology in the disclosure of new enemy of infective operators from higher plants. As of late, the utilization of plants as a wellspring of crucial mixes to battle microbial diseases has picked up noticeable quality. The need to look for plant-based antimicrobials is expanding because of significant expense, decreased adequacy and expanded protection from traditional drugs. This examination broke down the phytochemical structure of *Moringa oleifera*, and antimicrobial capability of its methanol and hexane extricates on *Escherichia coli*, *Klebsiella pneumonia*, *Pseudomonas aeruginosa* and *Candida albicans*. (Duraipandian et al. 2006; Mangale et al. 2012)

The investigation assessed antibacterial action of *Moringa oleifera* leaf concentrates and *Terminalia arjuna* bark separates against four microorganisms, viz. *Escherichia Coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Salmonella typhi*. Phytochemical investigation of the leaf in solvents of differing extremity; viz., methanol and chloroform were additionally done.

Phytochemical analysis of *Moringa oleifera* Linn has revealed a wide range of bioactive substances that enhance its medicinal and therapeutic value. Plant extracts derived through various techniques have been thoroughly examined for their phytochemical makeup, revealing a rich assortment of compounds such as alkaloids, flavonoids, phenolic compounds, and glycosides (Danjuma et al., 2025 ; Tambe & Gadhave, 2024). These substances are recognized for their antioxidant, antimicrobial, and potential therapeutic effects, establishing *Moringa oleifera* as a significant plant in both traditional and contemporary medicine. The following sections elaborate on the primary discoveries regarding the phytochemical composition of *M. oleifera*. *Moringa oleifera* extracts consistently contain alkaloids and flavonoids, which are known for their antioxidant and anti-inflammatory effects (Mali et al., 2024) (Kandeepan et al., 2022). Phenolic Compounds and Tannins are abundant in *Moringa oleifera* and contribute to their potent antioxidant activity, as shown by various assays such as DPPH and ABTS (Ogundipe et al., 2022) (Al-Reza et al., 2022). The presence of glycosides and terpenoids has been verified and linked to numerous health benefits, including anti-cancer and anti-thyroid activities (Mali et al., 2024) (Tambe & Gadhave, 2024). Extraction methods and various solvents such as ethanol, methanol, and ethyl acetate have been utilized to extract phytochemicals, with ethanol and ethyl acetate yielding higher concentrations of bioactive compounds (Shaeroun et al., 2019).

Based on the extraction method, different solvent was used which include ethanol and distilled water as well; ethanol usually results in descent concentration of all active compounds (Malhotra & Mandal, 2018). *Moringa oleifera* displays significant antioxidant and antimicrobial activities, attributed to its rich phytochemical content. These properties are advantageous for developing natural therapeutic agents (Seck et al., 2024). The Plant extracts have demonstrated potential in inhibiting enzymes, such as α -amylase and α -lipase, indicating their use in managing diabetes and obesity (Ogundipe et al., 2022). Although the phytochemical analysis of *Moringa oleifera* underscores its potential as a source of natural therapeutic agents, it is crucial to consider the variability in compound concentration due to different extraction methods and environmental factors. Further

research is necessary to standardize the extraction processes and to fully comprehend the pharmacological implications of these phytochemicals.

Moringa oleifera has high antibacterial activity against both Gram-positive (e.g. *Staphylococcus aureus*) and negative bacteria (e.g. *Escherichia coli*) (Danjuma et al., 2025) (Tambe & Gadhave, 2024) (Malhotra & Mandal, 2018). Zone of Inhibition: Substantial zones of inhibition mainly on ethanolic extracts with strong antibacterial effectiveness were reported (Souleymane et al., 2025).

The phytochemical attributes and antibacterial potential of *Terminalia arjuna* has been better documented, therefore, this herbal extract needs much more investigation for its full therapeutic benefit as only few resources are available regarding the use of *Moringa oleifera*.

On the other hand, there is a need for continuous examination of substitutes like *Terminalia arjuna* as the emergence of resistant strains means that *Moringa oleifera* cannot continue to be used successfully and replaces antibiotic which confirm their antibacterial activity.

The *Moringa oleifera* and *Terminalia arjuna* species show considerable potential for natural antimicrobials shown by qualitative phytochemical screening. *Moringa oleifera* leaves, in particular, have an impressive phytochemical profile and are effective against a variety of bacterial strains. The following sections provide significant insights on their phytochemical constituents and antibacterial potential. For phytochemical screening, some normal and accessible standard tests were finished. Antimicrobial bioassay was done through agar well dissemination technique.

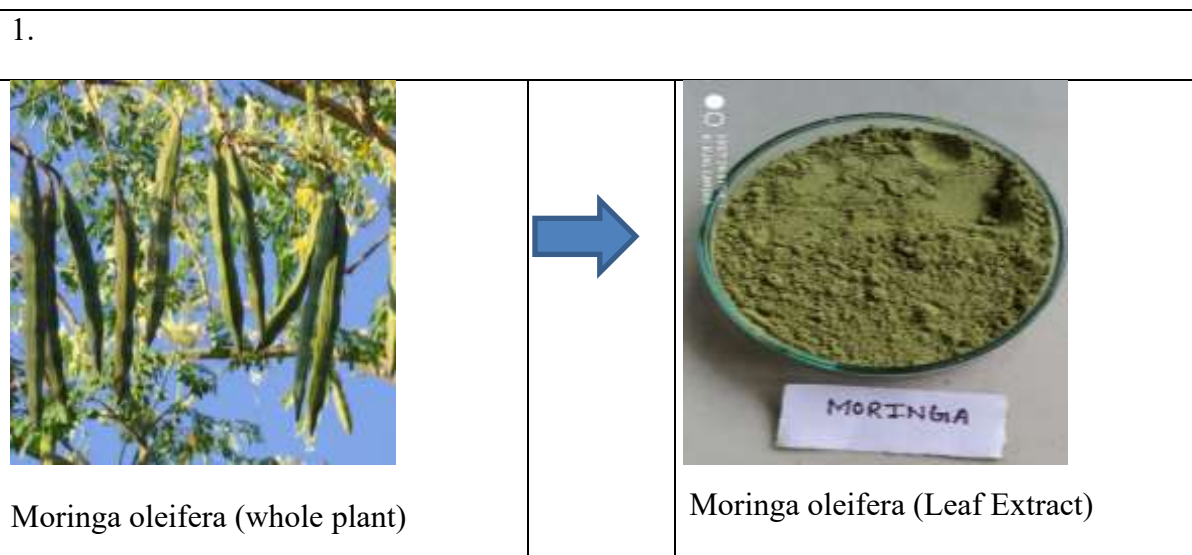
MATERIALS AND METHODS

Collection of the plant samples and preparation of the plant extracts of *Moringa oleifera* leaves and bark of *Terminalia Arjuna*.

Collection of Plant Materials:

Leaves of *Moringa* and bark of *Arjuna* were secured from nearby markets from Pune. It was guaranteed that the plant was solid and uninfected. The leaves were washed under running faucet water to dispense with dust then the plant materials were kept in until all the water content dissipated and the plant turned out to be all around dried for pounding. Subsequent to drying the plant material were ground utilizing with mechanical blender to get fine powder and the powder was put away in impermeable plastic holder with appropriate marking for sometime later.

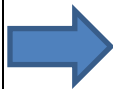
Identification



2.



Terminalia arjuna (whole plant)



Terminalia arjuna (Bark Extract)

Preparation of Leaf and Bark Extracts:

The dried leaf powder of *Moringa oleifera* and bark powder of *Terminalia arjuna* were extracted using different solvents, namely distilled water, methanol, ethanol, acetone, chloroform, and petroleum ether, following the maceration method. Briefly, 2 g of each powdered plant material was accurately weighed and transferred into separate 250 mL conical flasks. To each flask, 30 mL of the respective solvent was added. The flasks were kept at room temperature for 24 h with intermittent shaking to facilitate extraction of phytoconstituents.

After extraction, the mixtures were filtered through Whatman No. 1 filter paper. The filtrates were collected in clean Petri dishes and allowed to evaporate to dryness at room temperature for complete removal of the solvents. The dried extracts obtained were subsequently used for qualitative phytochemical screening.

Determination Of Extractive Value of Moringa Leaf and Arjuna’s Bark

The extractive value of each solvent extract was determined according to the method described by Khandelwal (2002). The filtrates obtained after extraction were concentrated to dryness by complete evaporation of the respective solvents. The dried extracts were weighed, and the percentage extractive value was calculated using the following formula:

$$\text{Extractive value (\%)} = \frac{\text{Weight of dried extract}}{\text{Weight of plant material}} \times 100$$

where:

- Weight of dried extract = weight of residue obtained after solvent evaporation (g)
- Weight of plant material = initial weight of powdered sample used for extraction (2 g)

The extractive values were determined for aqueous, methanolic, ethanolic, acetone, chloroform, and petroleum ether extracts of both plant materials.

Table I: Extractive value of *Moringa oleifera* (leaves):

Solvents	Weight of Plant material (g)	Colors of extract	Extractive value (%)
Aqueous (Water)	2	Green	5.00

Methanol	2	Green	14.50
Ethanol	2	Green	9.50
Acetone	2	Green	5.35
Chloroform	2	Green	6.30

Figure 1: Extractabilities (%) of active compounds by different solvents from *Moringa oleifera* (leaves)

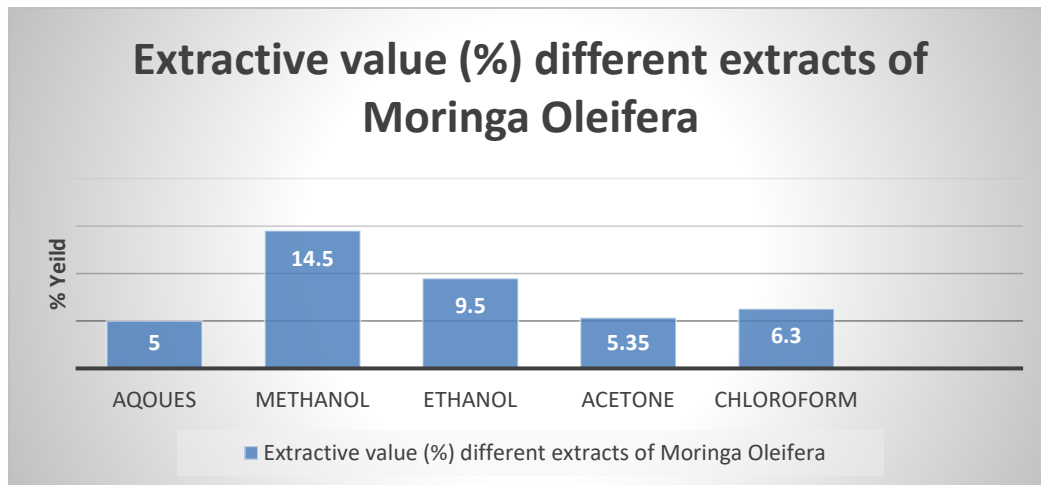
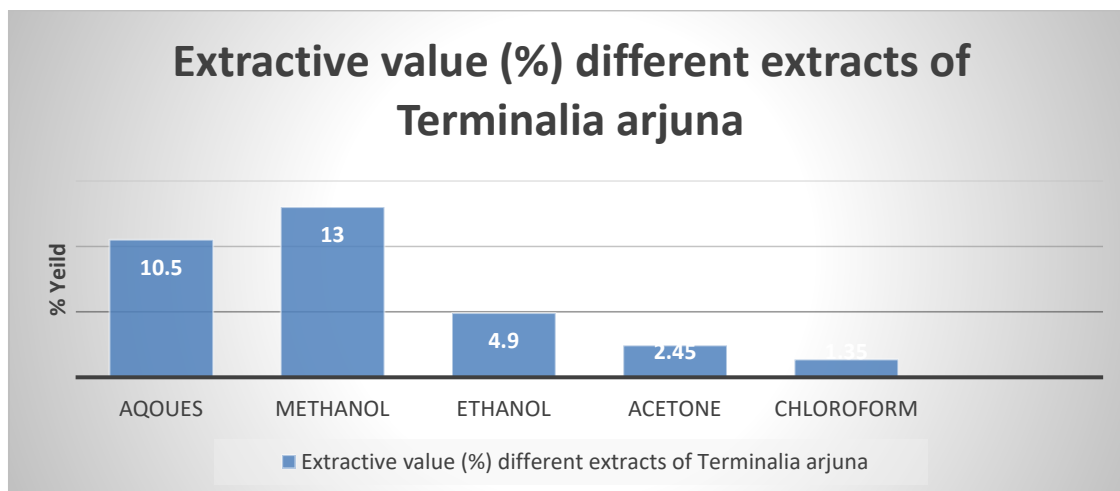


Table II: Extractive value of *Terminalia arjuna* (Bark)

Solvents	Weight of Plant material (g)	Colors of extract	Extractive value (%)
Aqueous (Water)	2	Brown	10.50
Methanol	2	Dark Brown	13.00
Ethanol	2	Dark Brown	4.90
Acetone	2	Yellowish	2.45
Chloroform	2	Yellowish	1.35

Figure 2: Extractabilities % of active compounds by different solvents from *Terminalia arjuna* (Bark)



5. Preliminary Phytochemical Analysis:

Phytochemical analysis (Rangari, V. D., 2002; Harborne, J. B., 1998) was done by the accompanying techniques to test for the nearness of the Phytosterol, Terpenoids, Total alkaloids, Carbohydrates, Flavonoids, Tannins, Proteins, Glycoside, Starch, Amino acids.

1. Test for Phytosterol:

Leibermann- Burchard reaction – To 3 ml extract, 10 ml chloroform was added followed by 2 ml of acetic anhydride. Then 2 drops of conc. Sulphuric acid was added from the side of the test tube. The blue green colour indicated the presence of steroids

2. Test for Terpenoids:

Salkowski test: The extract was mixed with 2 ml of chloroform and concentrated H₂SO₄ (3 ml) is carefully added to form a layer. A reddish-brown colouration of the interface is formed to show positive result of the presence of terpenoids. Salkowski test gave a positive result hence confirms the presence of Terpenoids

3. Test for Alkaloids

Mayer's reagent & Wagner's reagent confirmed the presence of Alkaloids in the extract. The Methanolic plant extract was warmed with 2% H₂SO₄ for two minutes. It is filtered and few drops of reagents were added separately.

- a. Mayer's reagent-A creamy- white colored precipitation appeared giving a positive result.
- b. Wagner's reagent-A reddish-brown precipitate appeared which also confirms the presence of alkaloids in the extract.

4. Test for Carbohydrate

Fehling's and Anthron's test confirmed the presence of carbohydrate. Fehling's Test: Fehling A and Fehling B reagents were mixed and few drops of extract was added and boiled. A brick red coloured precipitate of cuprous oxide forms, confirming the presence of carbohydrates

5. Test for Flavonoids

Ammonium Test: Plant extract heated with ethyl acetate for 3 min + mixture was filtered + 1ml ammonia solution was added. Layer's were allowed to separate. Yellow coloration at ammonia layer indicate the presence of flavonoid

6. Test for Tannins:

To 1.2 ml of extract of drug, added few drops of 5% FeCl₃ solution. A greenish colour indicates the presence of Galacto-tennins, while brown colour indicates Tannins

7. Test for Protein –

Millon's test - Dissolved small quantity of aqueous extract of drug in 1 ml of distilled water and 5-6 drops of millon's reagent. A white precipitate is formed which turns red on heating.

8. Test for Glycosides

Keller-Kiliani Test and Concentrate H₂SO₄ Test confirmed the presence of Glycosides in the methanolic plant extract. Keller-Kiliani Test: In 2 ml plant extract, glacial acetic acid, one drop of 5% FeCl₃ and conc.

H₂SO₄ were added. Reddish brown color appears at junction of the two liquid layers and upper layer appears bluish green, confirming the presence of glycosides. Concentrate H₂SO₄ Test: In 5 ml plant extract, 2 ml glacial acetic acid, one drop of 5% FeCl₃ and conc. H₂SO₄ were added. Brown ring appears, indicating the presence of glycosides.

9. Test for Starch

Dissolved 0.015 g of Iodine and 0.075 g of KI in 5 ml of distilled water and added 2-3 ml of an aqueous extract of drug. A blue colony is produced.

10. Test for amino acids

Ninhydrin test - About 3ml of plant extract solution was heated followed by addition of 3 drps of 5% ninhydrin solution. The test tubes with this solution were kept in boiling water bath for 10 minutes. The purple color was observed. It indicated the presence of amino acids.

Statistical Analysis

All experiments were performed in triplicate (n = 3), and results expressed as mean ± standard deviation (SD). Statistical analysis can be conducted using one-way Analysis of Variance (ANOVA) followed by Tukey's post hoc test to determine significant differences among solvent extracts. A probability value of p < 0.05 should be considered statistically significant.

Table III: Phytochemical screening of different solvent extracts of *Moringa oleifera*.

SR. NO.	Phyto-constituent	Phytochemical test	Results				
			WATER	MEOH	ETOH	CHLOROFORM	ACETONE
1	Phytosterols	Liebermann-Burchard's test	-	-	-	-	-
2	Terrpenoids	Salkowski reaction	+	-	+	+	+
3	Alkaloids	Mayer's test	-	-	-	-	-
		Wagner test	+	+	+	+	+
4	Carbohydrates	Fehling test	-	-	-	-	-
5	Flavanoids	Lead Acetate test	-	+	-	-	-
6	Tannins	5% FeCl ₃ Test	-	+	+	+	+
7	Proteins	Ninhydrin test	—	-	-	-	-
8	Glycosides	Keller-Killiani test	-	-	-	+	-
9	Starch	Iodine	-	-	-	-	-
10	Amino acid	Ninhydrine	+	-	-	-	-

Note: (+++) - High amount, (++) - Moderate amount, (+) - Less amount, (-) – Absent.

Table IV: Phytochemical screening of different solvent extracts of Terminalia arjuna

SR. NO.	Phyto-constituent	Phytochemical test	Results				
			WATER	MEOH	ETOH	CHLOROFORM	ACETONE
1	Phytosterols	Liebermann-Burchard's test	-	-	-	-	-
2	Terrpenoids	Salkowski reaction	+	-	+	+	+
3	Alkaloids	Mayer's test	-	-	-	-	-
		Wagner test	+	+	+	+	+
4	Carbohydrates	Fehling test	-	-	-	-	-
5	Flavanoids	Lead Acetate test	-	+	-	-	-
6	Tannins	5% FeCl ₃ Test	-	-	-	+	
7	Proteins	Ninhydrin test	—	-	-	-	-
8	Glycosides	Keller-Killiani test	-	-	-	+	-
9	Starch	Iodine	-	-	-	-	-
10	Amino acid	Ninhydrine	+	+	-	-	-

Note: (+++) - High amount, (++) - Moderate amount, (+) - Less amount, (-) – Absent.

Determination of the Antibacterial Activity: Selection of the Microorganisms

The bacterial strains utilized for this investigation were *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Salmonella typhii*. All the bacterial strains were developed and kept up in supplement agar. These living beings utilized were acquired from the Microbiology Department of MUIS of Ganpat University.

Well Diffusion Method:

The antibacterial movement of the leaf removes was resolved utilizing agar well dispersion strategy. The antibacterial movement of methanolic and chloroform concentrate of *Moringa oliefera* and *Terminilia arjuna* was tried on microbes viz. *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Salmonella typhii* by standard agar well dissemination technique. 1 ml organisms stock culture were vaccinated on supplement agar medium and filled sanitized Petri dishes. Wells of 5 mm distance across were made on the supplement agar utilizing a sterile plug borer. The cut agar plates were deliberately evacuated by the utilization of cleaned forceps. To each well, 20 µL of plant separates were stacked with the assistance of micropipette under aseptic conditions. The plates were containing development of the test living being and concentrates were hatched in well and plates were kept in hatchery at 37 °C for 24 h. The plates were analyzed for proof of zones of hindrance, which show up as a reasonable territory around the wells. The breadth of such zones of restraint was estimated. Control tests involving inoculums with gentamicin anti-toxin circles were arrangement and the plates were incubated at 37 °C for 24 h. The zones of restraint were then recorded and analyzed.

RESULT

The extractabilities of concentrates of *Moringa oleifera* and *Terminalia arjuna* was explored and described to in Table No. 1 and 2 From the current examination it was discovered that,. The highest extractive value obtained in methanol extract (14.5%) suggests that methanol efficiently extracts polar bioactive compounds from *M. oleifera* leaves. Similar observations were reported by Malhotra and Mandal (2018), who found methanolic extracts rich in flavonoids, tannins, and phenolic compounds. The presence of terpenoids, alkaloids, flavonoids, and tannins detected in the present study corroborates earlier reports by Danjuma et al. (2025) and Tambe and Gadhave (2024), indicating that *M. oleifera* is a valuable source of phytochemicals with therapeutic potential. The aqueous concentrates of *Moringa* and chloroform concentrate of *Arjuna* demonstrated lower extractive worth 5% and 1.35% than other dissolvable concentrates. The methanolic bark extract of *T. arjuna* showed the highest extractive value (13%), consistent with previous reports that methanol effectively extracts tannins, flavonoids, and triterpenoids from the bark. Nema et al. (2012) and Phani Kumar et al. (2013) similarly reported the presence of phenolic compounds and flavonoids contributing to antimicrobial activity. The observed antibacterial activity against *E. coli* and *Staphylococcus aureus* supports the traditional medicinal use of *T. arjuna* bark in treating microbial infections.

From the subjective examination of *Moringa oleifera* and *Terminalia arjuna* extracts the non-appearance of sterol, sugars, alkaloids, tannins, starch, amino corrosive were explored.

The different concentrates of *Moringa oleifera* in particular methanol and chloroform concentrates of its leaves and barks were tried against *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas vulgaris* for their antibacterial action. Compared with *M. oleifera*, *T. arjuna* exhibited stronger antibacterial activity, particularly against *Staphylococcus aureus* (11 mm inhibition zone). This difference may be attributed to the higher concentration of tannins, flavonoids, and triterpenoids present in the bark extracts. Similar findings have been reported in studies investigating the antimicrobial potential of *Terminalia* species.

Table V: Antibacterial activity of *Moringa oleifera*



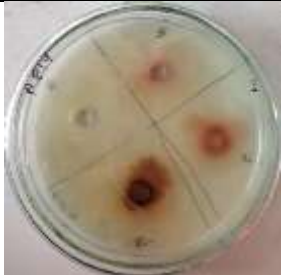



	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas vulgaris</i>
Chloroform			
Methanol			

Table VI: Antimicrobial activity of Chloroform, Methanol extract of medicinal plants against pathogens

Name of extract	Solvent	Zone of Inhibition (mm)		
		<i>Escherichia Coli</i>	<i>Pseudomonas vulgaris</i>	<i>Staphylococcus aureus</i>
<i>Moringa oleifera</i>	Methanol	-	-	-

	Chloroform	5	-	-
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Table VII: Antibacterial activity of Terminalia arjuna.





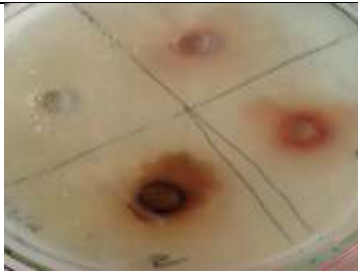

	Staphylococcus aureus	Escherichia coli	Pseudomonas vulgaris
Chloroform			
Methanol			

Table VIII: Antimicrobial activity of Chloroform, Methanol extract of medicinal plants against pathogens

Name of extract	Solvent	Zone of Inhibition (mm)		
		Escherichia Coli	Pseudomonas vulgaris	Staphylococcus aureus
Terminalia arjuna	Methanol	8	7	-
	Chloroform	6	-	11

DISCUSSION

Restorative plants are vital to the wellbeing of individual and networks (Pascaline et.al. 2011). Phytochemical examination led on the plant extracts uncovered the phytochemical constituents which are referred to display therapeutic activities (Sofowora, A., 1993). The phytochemical examination of Moringa oleifera plant removes utilizing water, methanol, chloroform, acetone, ethanol is appeared in Table no: 3 -From the phytochemical investigation it was discovered that sterol is missing in Moringa oleifera. Terpenoids were available in ethanol, fluid, chloroform, and acetone separate. Alkaloids were present Wagner test but remained undetected in Mayer's test (absolutely missing). Starch was also not detected. Flavonoids were available in methanol extract. Tannin was found in methanol, ethanol, chloroform, acetone separate however, protein was missing. Glycosides were found in chloroform separates but starch was thoroughly missing. Amino acids were found in the fluid concentrate. The phytochemical examination of Terminalia arjuna plant removes utilizing water, methanol, chloroform, acetone, ethanol is appeared in Table. From the phytochemical investigation it was discovered that sterol is missing. Terpenoids were available in ethanol, watery, chloroform, and acetone separate. Similar to the Moringa extracts, In T. arjuna extracts also showed Alkaloids were present in Wagner test but remained undetected in Mayer's test (absolutely missing). Starch was also not detected. Flavonoids were available in methanol concentrate of T. arjuna bark. Regular cancer prevention agents originate from plant as phenolic mixes, for example, flavonoids (Ali, 2008; Napoleonet.al. 2009). Tannin was found in chloroform remove. Protein was not found. Glycosides just found in chloroform extricate. Starch was thoroughly missing. Amino acids were just found in fluid and methanol separates.

The antibacterial action of chloroform, and methanol extricates was explored utilizing agar well dispersion strategy, against the chose human pathogens, for example, *Escherichia Coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*. All the inspected separate indicated fluctuating degrees of antibacterial exercises against the pathogens. Table-5 shows the antibacterial action of chloroform concentrate of *Moringa oleifera* indicated greatest zone of hindrance (5 mm) against *Escherichia Coli*. The antibacterial movement of chloroform concentrate of *Moringa oleifera* demonstrated no zone of restraint against *Pseudomonas aeruginosa*, *Staphylococcus aureus*. The antibacterial action of Methanol concentrate of *Moringa oleifera* indicated no zone of hindrance against the microbes on *Pseudomonas aeruginosa* (Karande et.al.2020; Deshpande et.al. 2022; Malhotra et.al. 2018). announced that *E.coli*, *P.aeruginosa*, *S. aureus* to be touchy to chloroform concentrate of *Moringa oleifera* leaf. Table-6 shows the antibacterial action of chloroform concentrate of *Terminalia arjuna* indicated most extreme zone of restraint (11 mm) against *Staphylococcus aureus*. The antibacterial action of chloroform concentrate of *Terminalia arjuna* demonstrated no zone of hindrance against *Pseudomonas aeruginosa* and, *Staphylococcus aureus*. The antibacterial action of methanol concentrate of *Terminalia arjuna* demonstrated greatest zone of restraint (8 mm) against *E.coli* and indicated the base inhibitory zone (6 mm) against *Pseudomonas aeruginosa*.

CONCLUSION

It can be concluded that the selected medicinal plants are the source of secondary metabolites like alkaloids, phytosterols, glycosides, phenols, flavonoids and terpenoids. Due to the presence of these secondary metabolites the selected medicinal plants have high healing potential.

Moringa oleifera and *Terminalia arjuna* show potential antimicrobial activity against tested bacteria viz, *E.coli*, *Pseudomonas aeruginosa* and *Staphylococcus aereus*. These two plants are considered in one of the new infection fighting strategies in controlling bacteria. These plants could be source of new antibiotic compound.

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